Tetrahedron Vol. 43. No. 2, pp. 383 to 389, 1987 Pnntcd I" Great **Bntam**

A FACILE SYNTHESIS OF ASCAMYCIN AND RELATED ANALOGUES

Julia Castro-Pichel, Maria Teresa Garcia-Lbpez* and Federico G. De las Heras Instituto de Química Médica. C.S.I.C., Juan de la Cierva 3, 28006-Madrid, Spain.

(Received in *UK 10 November 1986)*

Abstract - The nucleoside antibiotic ascamycin $|2$ -chloro-5'-0- $N-(L-alanyl)\sin famoyl$ adenosine (1)] has been synthesized by an improved procedure involving the direct condensation of 2-chloro-2',3'-0-isopropylidene-5'-0-sulfamoyladenosine (3) with Boc-L-Ala-OSu in DMF and in the presence of DBU. followed by removal of the protecting groups. A similar condensation of 3 with Boc-D-Ala-OSu and Boc-Gly-OSu, and subsequent deprotection, yielded the E-Ala and Gly analogues of 1, namely 2-chloro-5'-0-[N-CDalanyl)sulfamoyl]and 2-chloro-5'-0-[N-(glycyl)sulfamoyl]adenosine [D-ascamycin (14) and (18)]. Similar reactions of 2',3'-0-isopropylidene-5'-0-sulfamoyladenosine, (6) with the three amino acidderivatives above mentioned provided the corresponding adenosine analogues 12, 16 and 20. Several studies directed to demonstrate that the previous protection of the 6-NH₂ group of the adenosine derivatives 3 and 6 is not necessary for the selective aminoacylation of the SO₂NH₂ group are also reported.

Ascamycin (1) is a new nucleoside antibiotic recently isolated from a fermentation broth of Streptomyces¹. It shows a remarkably selective antibacterial activity as compared to dealanylascamycin [the antibiotic AT-265²,(2)³]. This selectivity in the case of <u>Xanthomonas</u> citri is due to a dealanylating enzyme present in the bacterial surface which, by removing the alanyl residue, affords dealanylascamycin and facilitates the selective uptake of this nucleoside derivative.

A synthesis of $\underline{\mathfrak{1}}$ has been reported by Isono et al 4,5 which involves condensation of $\underline{\mathtt{N}}^6$ -benzyloxycarbonyl (Z) or $\frac{\kappa^6}{2}$ -tert-butyloxycarbonyl(Boc)-2-chloro-2',3'-Q-isopropylidene-5'-Q-sulfamoyl

ÓМ

 $1.$ Ascamycin, $R = L -$ alanyl

 2 , Dealanylascamycin, $R = H$

- $3 \cdot R^{\dagger} = C1$; $R^2 = SO_2NH_2$
- **,R1=R2=li*
- 5 , R^1 = C₁; R^2 = H
- 2^2 . R^3 = H ; R^2 = SO₂NH₂

384 J. CASTRO-PICHEL et al.

adenosine with Z- or Boc-<u>L</u>-alanylimidazole in DMF using NaH as base, followed by removal of the protecting groups. While the present work was in preparation, a subsequent preliminary communication by the same group⁶ reported a related synthesis in which the aminoacylation step was catalyzed by cesium carbonate instead of NaH. According to this paper⁶, the use of NaH, employed in the first aminoacylations^{4,5}, resulted in considerable racemization of the alanine moiety (70% e.e.). The optical yield of the alanyl moiety using cesium carbonate was 88% e.e. No physical constants nor spectral data of any of the synthetic intermediates in the route to 1 were given in any of the above reports $4,5,6$.

As part of a project for the synthesis of $5'$ -0-sulfamoyl nucleosides as nucleotide analogues^{7,8,9}. we now describe an improved, facile synthesis of Ascamycin which consists on the direct condensation of <u>3</u>, without previous protection of the 6-NH₂ group, with Boc-L-Ala-OSu¹⁰ using 1.8-diazabicyclo [5, 4, 0] undec-7-ene (DBU) as base, to give 1, after deprotection, in a 76% yield from 3. This shorter procedure, in which no racemization of the L-Ala residue has been detected, has been extended to the synthesis of several amino acid modified analogues of 1 , including the D-Ala isomer, as well as to the corresponding adenosine analogues. The interest in the synthesis of $5'-0-[N-(amino-acy1)$ sulfamoyl]adenosine lies in the biological activity of $5'-0$ -sulfamoyladenosine itself $11,12,13$, which could be liberated after selective microbial deamino-acylation. This biological activity is similar to that of Nucleocidin (4'-fluoro-5'-O-sulfamoyladenosine) $^{11,14},$ which shows a broad antibacterial spectrum 15 and is particularly active against trypanosomes $^{16}.$

RESULTS AND DISCUSSION

The 2',3'-0-isopropylidene-5'-0-sulfamoyl derivatives of 2-chloroadenosine 3 and adenosine 6, were prepared in 80 and 71% yield, respectively, by reaction of 5 and 4 with bis(tri-n-butyltin) oxide and sulfamoyl chloride following Moffatt's procedure $^{11}.$

The key step in the preparation of 1 and analogues is the condensation of the 5'- 0 -sulfamoyl group of the corresponding 5'-0-sulfamoyladenosine derivative with the amino acid. Therefore, with the aim of attempting a direct condensation of the N^6 -unprotected derivatives 3 and 6, we compared the reactivities of the 5'-0-SO₂NH₂ and 6-NH₂ amino groups. In one of these previous experiments, the 5'-Q-sulfamoyladenosine derivative 6 was allowed to react with 1 equiv of N,N-dimethylformamide dimethylacetal¹⁷ giving, after 3 h, the corresponding 5'-0-[[M-(dimethylamino)methylene]sulfamoyl] derivative <u>7</u> in 85% yield as the only reaction product. The condensation of the 6-amino group of <u>6</u> with N,N-dimethylformamide dimethylacetal required 8 h and the use of an excess of the acetal.

Under these conditions the bis-[(dimethylamino)methylene]substituted nucleoside 8 was obtained. The attachment of the (dimethylamino)methylene function to the 5'-0-sulfamoyl moiety in 2 was deduced by the absence in its 1 H NMR spectrum of the peak at δ 7.55, attributed to the SO₂NH₂ protons of 6 and by the presence of the singlet at 6 7.30 assigned to the 6-NH₂ protons, as

compared to a similar signal δ 7.30 and 7.33 for the same protons of 6 and 5'-Q-sulfamoyl adenosine 13 respectively. These experiments suggested us the possible utilization of $\frac{N^6}{2}$ -unprotected-5'-0sulfamoyladenosine derivatives in the condensation with amino acids. Further support for this suggestion came out from the reaction of the 2-chloroadenosine derivative $\mathbf{5^{19}}$ with 1 equiv of Boc- $L = A1a-0$ Su in DMF and in the presence of Et $_3$ N (2 days), NaH (2h) or DBU (2h). These condensations $\frac{1}{1}$ led to the 5'-0-alanyl ester 9, while no traces of N° -alanyl derivatives were found. The ¹H NMR spectrum of 2 showed, besides the signals corresponding to the Ala moiety, the presence of the 6-NH₂ protons (6 7.80) and a downfield shift for the absorption of the 5'-CH₂ protons (6 4.23) as compared to the same protons $(6\ 3.70)$ of 5.

These results, prompted us to achieve the couplings of the N -hydroxysuccinimide esters of Boc-L-Ala, Boc-D-Ala, and Boc-Gly 10 with the 5'-O-sulfamoyladenosine derivatives $_2$ and $_2$, without previous protection of the 6-NH₂ group.

Condensation of $\frac{3}{2}$ with Boc-L-Ala-OSu in DMF using DBU as base gave 2-chloro-2',3'-Q-isopropylidene-5'-Q-[<u>N</u>-(Boc-L-alanyl)sulfamoyl]adenosine (<u>10</u>) in 80% yield which, after deprotection with trifluoroacetic acid (TFA)/H₂O (5:2), afforded 1^{20} . The structures of 10 and 1 were demostrated by analytical and spectroscopic data. Thus, the selective attachment of the alanyl moiety to the \texttt{suffix} sulfamoyl group in <u>10</u> was confirmed by the absence in its $^1\texttt{H}$ NMR spectrum of the singlet assigned to the SO₂NH₂ protons, which in 3 appeared at 67.53 and by the presence of a singlet at 67.80 , assigned to the 6-NH₂ protons, which disappeared by treatment with D_2O , as compared to a similar signal at δ 7.80 for the same protons of 3 and 5. Besides DBU, we have used Et₃N and NaH as bases, in the direct aminoacylation of $\frac{3}{5}$ with Boc-L-Ala-OSu. Although the best results, in terms of reaction time, clean work up and yield (45 and 75% using $Et_{\gamma}N$ and NaH respectively), were obtained with DBU, the exclusive aminoacylation of the $5'-0s0_{2}NH_{2}$ amino group was observed in all the cases.

Similar aminoacylation of 3 with Boc-D-Ala-OSu and Boc-Gly-OSu in the presence of DBU yielded the 5'-Q-[N-(Boc-aminoacyl)sulfamoyl]substituted nucleosides 13 and 17 in 80 and 71% yield respectively. Removal of the Boc and isopropylidene protecting groups of 13 and 17 with TFA/H₂0 (5:2) gave 2-chloro-5'-Q-[N-(Q-alanyl)sulfamoyl]adenosine[(14), Q-ascamycin] and 2-chloro-5'-Q-[(N-glycyl) sulfamoyl]adenosine (18) . Finally, a similar series of reactions of 6 with the N-hydroxysuccinimide esters of Boc-L-Ala, Boc-D-Ala, and Boc-Gly gave the corresponding 5'-O-[N-(Boc-aminoacyl)sulfamoyl] adenosine derivatives $\underline{11}$, $\underline{15}$ and $\underline{19}$ in 83, 80 and 74% yield respectively, which upon treatment with TFA/H₂0 (5:2) provided the corresponding deprotected compounds, namely 5'-0-[N-(L-alanyl), 5'-0-[N-(D-alanyl) and 5'-O-[N-(glycyl)sulfamoyl]adenosine 12, 16 and 20.

The structures of all these compounds ware determined from their 90- (for the protected derivatives 11, 13, 15, 17 and 19) or 360-MHz (for the unprotected compounds 12, 14, 16, 18 and 20) ¹H NMR

spectra. Thus, the attachment of the aminoacyl moiety to the sulfamoyl group in the protected 2 chloroadenosine derivatives 13 and 17 was determined, as in the case of 10, by the presence of singlets at δ 7.80 assigned to the 6-NH₂ protons and by the absence of the signal at 7.53 corresponding to the SO₂NH₂ protons of the starting sulfamoyl nucleoside 3. In a similar way, the structures of compounds 11 , 15 and 19 were established as $5'-Q-[\overline{N}-(aminoacy1)su1famoy1]$ substituted adenosine derivatives, based on their $¹$ H NMR spectra. They showed, in each case, a singlet at δ </sup> 7.28, assigned to the 6-NH₂ protons, as compared to the same protons of 4 and 6 which appeared at 6 7.32 and 7.30, respectively. However, the peak at 6 7.55 corresponding to the SO₂NH₂ protons of 6 did not appear in the spectra of 11 , 15 and 19 . As in the case of natural Ascamycin¹, the UV spectrum in H₂O of <u>1</u>, as well as those of the <u>D</u>-Ala and Gly analogs <u>14</u> and <u>18</u>,showed a maximum at 263 nm, while the UV spectra of 12 , 16 and 20 showed a maximum at 259 nm. These absorption maxima values correspond to those of the parent 5'-0-sulfamoyl nucleosides 3^{12} and $6^{11}.$ Modification of the chromophore by replacement of the 6-NH₂ group by acylamino or sulfonylamino produces a shift ~10 nm of the absorption maximum toward higher wawelength²¹. That no appreciable racemization of the alanyl moiety in compounds $1, 12, 14$ and 16 took place, was demonstrated by comparing the 360-MHz 1 H NMR spectra of the L-Ala substituted compounds 1 and 12 with those of the corresponding D -Ala isomers 14 and 16. This comparison showed that the 1 H NMR spectra of these couples of diastereoisomers (1, 14 and 12, 16) were not superimposable. It also showed that the spectrum of each **of** those compounds did not show detectable peaks of their corresponding diastereoisomer. Further evidence for these differences came out from the spectra of isomeric mixtures of 1 and 14 or 12 and 16 which in each case, clearly showed two different signals for the alanine CH_{3} protons.

In conclusion, the present method for the synthesis of Ascamycin is free of racemization and reduces the number of reaction steps in relation to those previously reported procedures^{4,5,6}. This improved method, involving the use of 6 N-unprotected sulfamoyl adenosine derivatives, allow to prepare analogs of Ascamycin in high yield.

EXPERIMENTAL

Melting points were measured with a Kofler hot-stage apparatus and are uncorrected. 'H NMR spectra were recorded with a Varian EM-390 and a Bruker HX-90-E spectrometer using Me_ASi as internal stardard. W absorption spectra were taken with a Perkin-Elmer 550 SE spectrophotometer. Optical rotations were determined with a Perkin Elmer 141 polarimeter. Analytical TLC was performed on aluminium sheets coated with a 0.2-mm layer of silica gel $60F_{254}$ purchased from Merck and preparative TLC on glass plates coated with a 2-mm layer of silica gel PF₂₅₄ (Merck). Silica gel 60(230-400 mesh) (Merck) was used for column chromatography.

2-Chloro-2',3'-O-isopropylidene-5'-O-sulfamoyladenosine (3). A suspension of 5^{19} (4.0 g, 11.4 mmol) in benzene (240 mL) containing hexabutyldistannoxane (13.8 g, 23 mmol) was refluxed under anhydrous conditions for 2 h. The resulting clear solution was cooled to 5°C in a dry box under nitrogen and a solution of sulfamoyl chloride (5.4 g, 47 mmol) in dioxane (80 mL) was added dropwise. After stirring for 30 min the solvent was removed and the residue was extracted with hot hexane. The insoluble residue was treated with dilute methanolic ammonia, evaporated and purified by chromatography on a silica gel columm using CHCl₃/MeOH(9:1)and then crystallized from acetone-CHCl₃ to give $\frac{3}{2}$ (3.9 g, 80%): m.p. 217-220°C (dec);[a] $\frac{20}{D}$ - 16.5° (c 1.0, MeOH); 1 H NMR (DMSO-d₆) 6 1.30 and 1.50 (Zs, 6H, isopropylidene), 4.15(m, ZH, H-5'), 4.36(m, lH, H-4'), 4.98(dd, lH, H-3'), 5.32(dd, lH, H-2'), 6.13(d, 1H, H-1', J_{1',2'}= 2 Hz), 7.53(br s, 2H, SO₂NH₂, D₂O exchangeable), 7.80 (br s, 2H, NH₂-6, D₂0 exchangeable), 8.27 (s, 1H, H-8). Anal. Calcd. for $C_{13}H_{17}C1N_6O_6S$: C, 36.70; H, 4.00; Cl, 8.35; N, 19.76; S, 7.53. Found: C, 36.86; H, 4.29; Cl, 8.46; N, 19.39; S, 7.63.

 $2', 3'-0$ -Isopropylidene-5'-0-sulfamoyladenosine (6). $2', 3'-0$ -Isopropylideneadenosine (4, 3.0 g, 9.7 mmol) was treated with hexabutyldistannoxane (11.5 g, 20 mmol) and sulfamoyl chloride (4.5 g, 39 mmol) as described for the preparation of 3. Purification by column chromatography using CHCl₃-MeOH (9:1) and crystallization from acetone-chloroform gave 6 (2.70 g, 71%): m.p. 205-207°C (dec);[a] $\frac{20}{0}$ -23.2° (c_1.0, MeOH); ¹H NMR (DMSO-d_c) 6 1.32 and 1.56 (2s, 6H, isopropylidene), 4.20(m, 2H, H-5'),

4.36(m, lH, H-4'), 5.08(dd, lH, H-3'), 5.43(dd, lH, H-Z'), **6.22(d, 1H, H-l', J1, 2,= 2 HZ),** 7.30 (br s, 2 H, NH₂-6, D₂0 exchangeable), 7.55 (br s, 2H, SO₂NH₂, D₂0 exchangeable), 8.16(s, 1H, H-2), 8.30(s, 1H, H-8). Anal. Calcd. for $C_{1,3}H_{1,8}N_{6}O_6S$: C, 40.40; H, 4.66; N, 21.76; S, 8.39. Found: C, 40.76; H, 4.52: N, 21.53; S. 8.31.

Reaction of 6 with N,N-dimethylformamide dimethyl acetal. To a solution of 6 (0.1g, 0.24 mmol) **in DMF** (2 mL) cooled at O°C was added dropwise N,N-dimethylformamide dimethyl acetal (0.03 g, 0.25 mmol) in DMF (1 mL) and the resulting mixture was stirred for 3 h. Then the solvent was evaporated to dryness, the residue was dissolved in CHCl₃ (1 mL) and the solution was added, with stirring, to petroleum ether (20 mL). The resulting precipitate was collected and dried over P_0O_F to give pure 2',3'-0-isopropylidene-5'-O-[[N-(dimethylamino)methylene] sulfamoylladenosine (7_) (0.11 g, 85%): m.p. 176 -178°C; ¹H NMR (DMSO-d₆) & 1.32 and 1.54(2s, 6H, isopropylidene), 2.83 and 3.10 [2s, 6H, N(CH3)2], 4.10(m, 2H, H-5'), 4.36(m, lH, H-4'), 5.05(dd, lH, H-3'), 5.42(dd, lH, **H-2'), 6.20(d,** lH, H-1', J_{1, 2},= 2 Hz), 7.30(br s, 2H, NH₂-6, D₂0 exchangeable), 8.06[s, 1H, CH-N(CH₃)₂], 8.15(s, 1H, H-2), 8.28(s, 1H, H-8). Anal. Calcd. for $C_{16}H_{23}N_70_6S$: C, 44.96; H, 5.60; N, 22.22; S, 7.49. Found: C, 44.62; H, 5.46; N, 22.36; S, 7.37.

To a solution of 2 (0.10 g, 0.24 **mmol) in** DMF (2 mL) was added N,N-dimethylformamide dimethyl acetal (0.10 g, 0.8 mmol) in DMF (2 mL) and the mixture was kept at room temperature for 8 h. Then the solvent was removed and the residue was treated as above to provide 2^1 ,3'-O-isopropylidene-N^C-[(dimethylamino)methylene]-5'-0-[[N-(dimethylamino)methylene]sulfamoyl]adenosine (8) (1.09 g, 84%): m.p. 181-183°C; 1 H NMR (DMSO-d₆) 6 1.32 and 1.54(2s, 6H, isopropylidene group), 2.83, 3.10, 3.15 and 3.20 [4s, 12H, NCH_3], 4.13(m, 2H, H-5'), 4.40(m, 1H, H-4'), 5.07(dd, 1H, H-3'), 5.48(dd, 1H, H-2'), 6.30(d, 1H, H-1', $J_{1',2'}$ = 2 Hz), 8.06 and 8.92 [2s, 2H, CH-N(CH₃)₂], 8.35(s, 1H, H-2), 8.40(s, 1H, H-8). Anal.Calcd. for $C_{19}H_{28}N_8O_6S$: C, 45.96; H, 5.64; N, 22.58; S, 6.45. Found: C, 45.58; H, 5.80; N, 22.23: S, 6.73.

Reaction of 5 with Boc-L-Ala-OSu. A stirred solution of 5 (0.2 g, 0.57 mmol) and Boc-L-Ala-OSu (0.17 g, 0.58 mmol) in DMF (4 mL) was kept at room temperature in the presence of 1 equiv of the following bases and for the following reaction times: a) triethylamine for two days, b) sodium hydride or DBU for 2 h. Evaporation of the solvent left a residue which was purified by preparative TLC using CHCl₃-MeOH (9:1) to afford 5'-O-(Boc-L-alanyl)-2-chloro-2',3'-O-isopropylideneadenosine 9 [a) 0.10 g, 34%, b) 0.17 g, 60%]: m.p. 129-131°C(from MeOH-ether); ¹H NMR (DMSO-d_c) 61.16 (d, 3H, Ala $f{B}$ CH₃), 1.30 and 1.50(3s, 15H, Boc and isopropylidene groups), 3.82(m, 1H, Ala a CH), 4.23 (m, 3H, H-4' and H-5'), 5.02 (dd, 1H, H-3'), 5.38(dd, 1H, H-2'), 6.13(d, 1H, H-1', J_{1', 2},= 2 Hz), 7.20(d, 1H, Ala NH, D₂0 exchangeable), 7.80(s, 2H, NH₂-6, D₂0 exchangeable), 8.30(s, 1H, H-8). Anal. Calcd. for $C_{21}H_{20}C1N_6O_7$: C, 49.17; H, 5.66; Cl, 6.93; N, 16.39. Found: C, 48.97; H, 5.84; Cl. 6.57; N, 16.04.

General procedure for the condensation of the 5'-0-sulfamoyl nucleosides 3 or 6 with N-Boc-L-Ala-OSu, N-Boc-D-Ala-OSu and N-Boc-Gly-OSu. A solution of the 5'-O-sulfamoyl nucleoside (1 mmol) and DBU (1 mmol) in DMF (6 mL) was treated with the N-hydroxysuccinimide ester of the Boc-amino acid (1 mmol) and the mixture was stirred at room temperature for 1.5 h. After evaporation of the solvent the residue was purified by column chromatography eluting with CHC1₃-MeOH (9:1) to provide the corresponding 5'-^o-[N-(Boc-aminoacyl)sulfamoyl]nucleosides which were identified as specified in each case.

2-Chloro-2',3'-0-isopropy1idene-5'-O-[N-(Boc-L-alanyl)sulfamoyl]adenosine (lo). 80% Yield: m.p. > 250°C (from MeOH-ether)[a] $_D^{20}$ -23° (c 0.5 MeOH); 1 H NMR (DMSO-d₆) 6 1.16(d, 3H, Ala BCH₃); 1.32 and 1.50(2s, 6H, lsopropylidene group), 1.36(s. 9H, Boc), 3.68(m, lH, AlaaCH), 4.02(m, 2H. H-5'), 4.40 (m, 1H, H<mark>-4'), 4.93(dd, 1H, H-3'), 5.25(dd, 1H, H-2'), 6.08(d, 1H, H-1', J_{1',2'}= 2.5 Hz), 6.33(d,</mark> 1H, Ala NH, D₂0 exchangeable), 7.80(s, 2H, NH₂-6, D₂0 exchangeable), 8.40(s, 1H, H-8). Anal. Calcd. for $C_{21}H_{30}C1N_7O_9S: C$, 42.60; H, 5.07; C1, 6.00; N, 16.56; S, 5.40. Found: C, 42.37; H, 4.76; C1, 6.39; N, 16.27; S, 5.45.

 2^1 ,3'-O-Isopropylidene-5'-O-[N-(Boc-L-alanyl)sulfamoyl]adenosine (11). 83% Yield: m.p. 211-213°C (dec) (from MeOH-ether); $[\alpha]_D^{20}$ - 32.8° (c 0.5, MeOH); ¹H NMR (DMSO-d₆)6 1.16(d, 3H, Ala BCH₃), 1.30 and 1.55(2s, 6H, isopropylidene group), 1.34(s, 9H, Boc), 3.68(m, 1H, Ala aCH), 4.00(m, 2H, H-5'), 4.36(m, 1H, H-4'), 5.00(dd, 1H, H-3'), 5.32(dd, 1H, H-2'), 6.16(d, 1H, H-1', $J_{1',2'}$ = 2 Hz), 7.28

(s, 2H, NH₂-6, D₂O exchangeable), 8.14(s, 1H, H-2), 8.36(s, 1H, H-8). Anal. Calcd. for C₂₁H₃₁N₇O₉S: C, 45.24; H, 5.56; N, 17.60; S, 5.75. Found: C, 44.88; H, 5.27; N, 17.35; S, 5.46.

2-Chloro-2',3'-O-isopropylidene-5'-O- [N-(<u>Boc-D-alanyl)sulfamoyl]adenosine</u> (13). 80% Yield: $m.p.$ > 250°C (from MeOH-ether); [α] $\frac{20}{D}$ - 15.6° (c 0.5, MeOH); ¹H NMR (DMSO-d₆)6 1.18(d, 3H, Ala B CH₃); 1.32 and 1.50 (2s, 6H, isopropylidene group), 1.36(s, 9H, Boc), 3.68(m, lH, AlaoCH), 4.OO(m, 2H, H-5'), 4.40(m, 1H, H-4'), 4.95(dd, 1H, H-3'), 5.25(dd, 1H, H-2'), 6.10(d, 1H, H-1', $J_{11,21}$ = 2 Hz), 7.80(s, 2H, NH₂-6, D₂0 exchangeable), 8.40(s, 1H, H-8). Anal. Calcd. for $C_{21}H_{30}C1N_7O_qS$: C, 42.60; H, 5.07; Cl, 6.00; N, 16.56; S, 5.40. Found: C, 42.41; H, 4.78; Cl, 6.37; N, 16.42; S, 5.43.

2',3'-O-Isopropylidene-5'-O-[N-(Boc-D-alanyl)sulfamoyl]adenosine (15). 80% Yield: m.p. 209-210°C (dec) (from MeOH-ether);[a] $^{20}_{D}$ - 4.3° (c 0.5, MeOH); ¹H NMR (DMSO-d₆) 6 1.15(d, 3H, Ala B CH₃), 1.30 and 1.55(2s, 6H. isopropylidene group), 1.33(s, 9H, Boc), 3.68(m, lH, *Ala 0* CH), 4.00(m, 2H, H-5'), 4.36(m, 1H, H-4'), 5.00(dd, 1H, H-3'), 5.35(dd, 1H, H-2'), 6.16(d, 1H, H-1', J_{1',2'}= 2 Hz), 7.28 (s, 2H, NH₂-6, D₂0 exchangeable), 8.12(s, 1H, H-2), 8.35(s, 1H, H-8). Anal. Caldc. for C₂₁H₃₁ ~~0~s: c, 45.24; H, 5.56; N, 17.60; S, 5.75. Found: C, 44.87; H, 5.21; N, 17.39; S, 5.72.

2-Chloro-2',3'-0-isopropylidene-5'-0-[N-(Boc-glycyl)sulfamoyladenosine (17). 71% Yield: m.p.> 250°C (from MeOH-ether); $[a]_D^{20}$ - 10.8° (c, 0.5, MeOH); ¹H NMR (DMSO-d₆) 6 1.32 and 1.50 (2s, 6H, isopropylidene group), $1.35(s, 9H, Boc)$, $3.40(d, 2H, Gly CH_2)$, $4.00(m, 2H, H-5')$, $4.40(m, 1H, H-4')$, 4.95(dd, 1H, H-3'), 5.25(dd, 1H, H-2'), 6.10(d, 1H, H-1', $J_{1',2'}$ = 2 Hz), 7.80(s, 2H, NH₂-6, D₂0 exchangeable), 8.40(s, 1H, H-8). Anal. Calcd. for $C_{20}H_{28}C1N_7O_9S: C$, 41.56; H, 4.84; Cl, 6.15; N, 16.97; S, 5.54. Found: C, 41.18; H, 4.46; Cl, 6.38; N, 16.71; S, 5.45.

 $2^{\prime},3^{\prime}-0$ -Isopropylidene-5'-O-(N- $Boc-glycy1/sulfamoy1]$ adenosine (19). 74% Yield: m.p. 208-210 $^{\circ}$ C(dec) (from MeOH-ether); $[a]_D^{20}$ - 16.1° (c 0.5, MeOH); ¹H NMR (DMSO-d₆)61.30 and 1.50 (2s, 6H, isopropylidene group), 1.34(s, 9H, Boc), 3.37(d, 2H, Gly CH₂), 4.00(m, 2H, H-5'), 4.35(m, 1H, H-4'), 5.00(dd, 1H, H-3'), 5.35(dd, 1H, H-2'), 6.15(d, 1H, H-1', $J_{1',2'}$ = 2 Hz), 7.28(s, 2H, NH₂-6, D₂0 exchangeable), 8.13(s, 1H, H-2), 8.35(s, 1H, H-8). Anal. Calcd. for $C_{20}H_{29}N_7O_9S$: C, 44.20; H, 5.34; N, 18.05; S, 5.89. Found: C, 43.89; H, 5.00; N, 17.78; S, 5.76.

Deprotection reactions. General procedure. A suspension of the corresponding 2',3'-O-isopropylidene-5'-0-[N-(Boc-aminoacyl)sulfamoyl]nucleoside (0.5 mmol) in a mixture of TFA/H₂0 (5:2) (4 mL) was stirred at room temperature for 2 h. The solvent was evaporated and the residue was coevaporated three times with EtOH and purified by column chromatography, eluting with CHCl₃-MeOH (2:1) for compounds 1, 12, 14 and 16, and with CHCl₃-MeOH (1:1) for compounds 18 and 20.

2-Chloro-5'-0-[N-(L-alanyl)sulfamoyl]adenosine (Ascamycin, 1). 95% Yield: m.p.>270°C (from EtOH-H₂O), lit^{1,2O} m.p. >270°C;[a]^{2O} -3.7°(c 0.5, H₂O), lit^{1,2O} [a]+2.3°(c 1.0, H₂O); UV λ_{max} (H₂O) 263 nm (ε 1250O); lit^t UV x_{max} (H₂O) 263 nm (ε 1227O); ⁻H NMR (D₂O, 36O MHz) 6 1.26(d, 3H, Ala B CH₃, J= 7.18 Hz), 3.60(q, 1H, Ala a CH), 4.21(m, 2H, H-5'), 4.25(m, 1H, H-4'), 4.31(t, 1H, H-3'), 4.52(t, 1H, H-2'), 5.83(d, 1H, H-1', J_{1',2'}= 4.70 Hz), 8.14(s, 1H, H-8). Anal. Calcd. for $C_{13}H_{18}C1N_2O_7S: C, 34.55; H, 3.99; Cl, 7.86; N, 21.70; S, 7.08. Found: C, 34.88; H, 4.32; Cl, 7.58;$ N, 21.46; S, 6.92.

5'-O-[N-(L-Alanyl)sulfamoyl]adenosine (12). 94% Yield: m.p. 215°C (dec) (from EtOH-H₂O); $[a]_n^{-2}$ – 12.8°(c 0.5, H₂0); UV λ_{max} (H₂0) 259 nm (e 11300); "H NMR (DMSO-d_e+D₂0, 360 MHz) 6 1.30(d, 3H, Ala BCH₃, J = 7.14 Hz), 4.05-4.17(m, 4H, H-3', H-4', H-5'), 4.58(t, 1H, H-2'), 5.91(d, 1H, H-1', $J_{1',2'}$ = 5.86 Hz), 8.14(s, 1H, H-2), 8.37(s, 1H, H-8). Anal. Calcd. for $C_{1,3}H_{1,9}N_7O_7S$: C, 34.51; H, 4.20; N, 21.68; S, 7.08. Found: C, 34.32; H, 4.50; N, 21.53; S, 7.00.

2-Chloro-5'-O-[N-(D-alanyl)sulfamoyl)]adenosine (D-Ascamycin, 14). 95% Yield: m.p.> 250°C (from EtOH-H₂O);[a] $_{D}^{20}$ -11.5°(c 0.5, H₂O); UV $_{max}$ (H₂O) 263 nm (ϵ 13100); ¹H NMR (D₂O, 360 MHz) 6 1.27(d, 3H, Ala β CH₃, J = 7.18 Hz), 3.64(q, 1H, Ala α CH), 4.20(m, 2H, H-5'), 4.23(m, 1H, H-4'), 4.31(t, 1H, H-3'), 4.52(t, 1H, H-2'), 5.83(d, 1H, H-1', $H_{1',2'}$ = 4.92 Hz), 8.14(s, 1H, H-8). Anal. Calcd. for C₁₃H₁₈ClN₇0₇S: C, 34.55; H, 3.99; Cl, 7.86; N, 21.70; S, 7.08. *Found: C, 34.43*; H, 3.66; Cl, 7.63; N, 21.53; S, 6.82.

5'-O-[N-(D-Alanyl)sulfamoyl]adenosine (16). 93% Yield: m.p. 212°C (dec);[a] $^{20}_{D}$ -18.3°(c 0.5, H₂0); UV λ _{max} (H₂O) 259 nm (e 13000); 1 H NMR (DMSO-d₆ +D₂O, 360 MHz)6 1.28(d, 3H, Ala B CH₃, J=

7.10 Hz), 4.03-4.18(m, 4H, H-3', H-4', H-5'), 4.59(t, 1H, H-2'), 5.91(d, 1H, H-1', $J_{11,21}$ = 5.87 Hz), 8.14(s, 1H, H-2), 8.37(s, 1H, H-8). Anal. Calcd. for $C_{13}H_{10}N_7O_7S$: C, 34.51; H, 4.20; N, 21.68; S, 7.0%. Found: C, 34.21; H, 4.45; N, 21.34; S, 7.41.

2-Chloro-5'-0[N-(Clycyl)sulfamoyl]adenosine (18). 96% Yield: m.p.> 250°C; [o] $^{20}_{0}$ -15.4° (c 0.5, $H_2(0)$; UV λ_{max} (H₂0) 263 nm (ϵ 12200); ¹H NMR (DMSO-d₆+D₂0, 360 MHz)6 3.24(s, 2H, Gly CH₂), 4.05-4.15 (m, 4H, H-3', H-4', H-5'), 4.50(t, 1H, H-2'), 5.83(d, 1H, H-1', $J_{1',2'}$ = 5.96 Hz), 8:41(s, 1H, H-8). Anal. Calcd. for $C_{12}H_{16}C1N_7O_7S$: C, 32.88; H, 3.65; Cl, 8.10; N, 22.37; S, 7.31. Found: C, 32.50; H, 4.03: Cl, 8.28; N, 22.68; S, 7.65.

 $5'-0-[N-(Glycy1)su1famoy1]$ adenosine (20). 95% Yield: m.p. 212°C (dec);[a] $^{20}_{D}$ - 22.3° (c 0.5, $H_2(0)$; UV λ_{max} (H₂0) 259 nm (ϵ 11000); ¹H NMR (DMSO-d₆+D₂0, 360 MHz) δ 3.22(s, 2H, Gly CH₂), 4.04-4.15(m, 4H, H-3', H-4', H-5'), 4.58(t, 1H, H-2'), 5.90(d, 1H, H-1', $J_{1^4,2^7}$ 5.86 Hz), 8.14(s, 1H, H-2), 8.38(s, 1H, H-8). Anal. Calcd. for $C_{12}H_{17}N_7O_7S$: C, 35.62; H, 3.88; N, 22.37; S, 7.30. Found: C, 35.71; H, 4.25; N, 22.27; S, 7.02.

ACKNOWLEDGMENTS. We thank the Comisión Asesora de Investigación Científica y Técnica for financial support and the Ministerio de Asuntos Exteriores for a fellowship. We also thank Dr. J. L. Nieto (Instituto de Estructura de la Materia, C.S.I.C.) for recording the 1 H NMR spectra at 360 MHz.

REFERENCES

- 1. K. Isono, M. Uramoto, H. Kusakabe, N. Miyata, T. Koyama, M. Ubukata, S.K. Sethi, and 3.A. McCloskey, J. Antibiot., 37, 670 (1984).
- 2. E. Takahashi, and T. Beppu, J. Antibiot., 35, 939 (1982).
- 3. H. Osada, and K. Isono, Antimicrob. Agents Chemother., 27, 230 (1985).
- 4. K. Isono, *M.* Uramoto, H. Osada, M. Ubukata, H. Kusakabe, T, Koyama, N. Miyata, S.K. Sethi, and J.A. McCloskey, Nucleic Acids Research, Symposium Series, 15, 65 (1984).
- 5. *M. Ubukata, H. Osada, and K. Isono, Nucleic Acida Research, Symposium Series, 16, 81 (1985).*
- 6. M. Ubukata, and K. Isono, <u>Tetrahedron Lett., 27, 3907 (198</u>6).
- 7. M.J. Camarasa, P. Fernández-Resa, M.T. García-López, F.G. De las Heras, P.P. Méndez-Castrillón, B. Alarcón, and L. Carrasco, J. Med. Chem., 28, 40 (1985).
- 8. *P.* Fern&ndez-Resa, M.T. Garcia-Lopez, F.G. De las Heras, A. San Felix, 8. Alar&n, and L. Carrasco, <u>Eur. J. Med. Chem</u>., <u>21</u>, 245 (1986).
- 9. J. Andrés, M.T. García-López, F.G. De las Heras and P.P. Méndez-Castrillón, Nucleosides Nucleotides, In press.
- 10. G.W. Anderson, J.E. Zimmerman, and F.M. Callahan, J. Am. Chem. Sot., 86, 1839 (1964). I.D. Jenkins, J.P.H. Verheyden , and 3.G. Moffatt, J. **Am, Chem. Sac., 98, 3346 (1976).**
- 11.
- 12. G.R. Gough, D.M. Nobbs, F. Penglis-Caredes, and M.H. Maguire, J. Med. Chem., 21, 520 (1978).
- 13. D.A. Shuman, M.J. Robins, and R.K. Robins, J. *Am.* Chem. Sot., 92, 3434 (1970).
- 14. 15. A. Bloch, and 6. Coutsogeorgopoulos, Biochemistry, lo, 4394 (1971). S.O. Thomas, V.L. Singleton, J.A. Lowery, R.W. Sharpe, L.M. Pruess, J.N. Porter, J.H. Mowat,
and N. Bohonos, <u>Antibiot. Annu., 7</u>, 716 (1956–1957).
- 16. R.I. Hewitt, A.R. Gumble, L.H. Taylor, and W.S. Wallace, Antibiot. Annu., 2, 722 (1956-1957),
- 17. 18. 2. Arnold, and M. Kornilov, Collect. Czech. Chem. Commun., 29, 645 (1964).
- 19. L.C. March, and K.C. Taou, J. Heterocyclic Chem., 2, 685 (1970).
- 20. G.R. Gough, M-H. Maguire, and F. Michal, J, Med. Chem., 12, 494 (1969). Attempts to get a sample of natural Ascamycin were unsuccessful. Thus, comparison of (1) with the natural antibiotic was not possible.
- 21. R.S. CunnIngham and M.U, Gray, Blochemtstry, 13, 543 (1974).